

SYNTHESIS OF SOME NEW THEOPHYLLINE DERIVATIVES USING CLICK CHEMISTRY AND STUDY OF BIOLOGICAL ACTIVITY

Ruaa J. Kamel¹, Nabeel A. Abdul-Rida^{2*} 

¹Department of pharmaceutical chemistry, College of pharmacy, Al-Qadisiyah University, Iraq

²Department of Chemistry, College of science, Al-Qadisiyah University, Iraq

*email: nabeel.a.alradha@qu.edu.iq

Received 07.11.2024

Accepted 13.01.2025

Abstract: In this study, the authors chemically synthesized new theophylline derivatives and then evaluated them in vitro as antimicrobial agents. The target derivatives, theophylline derivatives conjugated with 1,2,3-triazole and piperazine moieties, were synthesized via sequential various reactions. Spectroscopically, the researchers employed FT-IR, ¹H NMR, and ¹³C NMR techniques to get further insights into these derivatives; they also used the CHN technique to facilitate their structure identification. In addition, the authors conducted tests in vitro to evaluate the antibacterial activity of all the synthesized derivatives against *E. coli* and *S. aureus*. The study results obtained showed encouraging antibacterial efficacy.

Keywords: click reaction, antibacterial agent, theophylline, triazole derivatives.

DOI: 10.32737/2221-8688-2025-4-549-555

Introduction

Copper-catalyzed alkyne-azide cycloaddition (CuAAC) reactions are known as "click" chemistry reactions, where they produce 1,2,3-triazoles. Click reactions' high reaction yield, broad substrate range, and ease of purification make them one of the most advantageous catalytic processes for the synthesis of 1,2,3-triazoles [1-4]. Simple copper salts such as CuI, Cu(AcO)₂, and CuSO₄ are preferred as catalysts in the bulk of published protocols for these reactions [5-10]. For more than 80 years, methylxanthines, in particular theophylline, have been known to be effective bronchodilators for the treatment of acute asthma [11]. Theophylline (Fig. 1A), referred to as 1,3-dimethylxanthine, belongs to the purine class; it is a well-established bronchodilator medication that has been used to treat respiratory diseases like asthma and chronic pulmonary obstructive disease (COPD) [12-14]. Adenosine receptor antagonism is one of theophylline's modes of action [15]. Cyclic AMP and cyclic GMP intracellular concentrations rise as a result of theophylline's poor and non-selective inhibition of

phosphodiesterases (PDEs) of cyclic nucleotides [16]. The drug's strong antagonistic action at adenosine receptors is its other well-documented pharmacological function [17]. Additionally, theophylline has been demonstrated to increase the release of interleukin-10 (likely through the inhibition of PDE), antagonize tumor necrosis factor alpha, inhibit the pro-inflammatory effects of prostaglandins, decrease intracellular calcium ion release, prevent nuclear factor-κB from translocating into the nucleus (which may lower the expression of inflammatory genes), stimulate apoptosis, and increase histone deacetylase activity [18-20]. In addition, theophylline and some of its derivatives (Fig. 1) are important heterocyclic scaffolds for the development of effective pharmacological treatments, including antimicrobial [21], antiviral (for HCV and COVID-19) [22, 23], antituberculosis properties [24], anti-cancer [25], and anti-inflammatory effects [26]. In the present study, we synthesize new derivatives of 1,2,3-triazole in addition to their biological activity evaluation.

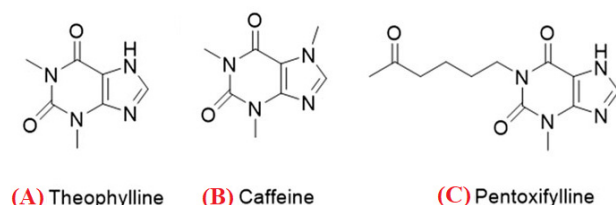


Fig. 1. Theophylline and some of its derivatives

Experimental part

Chemicals and Analytical Devices. The materials were provided by Sigma-Aldrich (USA), BDH (United Kingdom), and Merck (Germany). The measurement was obtained using a melting-point thermometer. A thin-layer chromatography (TLC) technique was employed to monitor the progress of the reaction using a 9:1 mixture of benzene and methanol as the eluent. After completion, the solvent had completely evaporated. The purification of the products was carried out using column chromatography. The column had a diameter of 2 cm and a length of 50 cm. Silica gel with a mesh size of 60-120 was used as the stationary phase. Different eluents were employed based on the separation observed on thin-layer chromatography (TLC). The spectra were obtained using a Shimadzu FT-IR (8300) spectrophotometer equipped with KBr discs in the range of 4000-400 cm^{-1} . The ^{13}C NMR and ^1H NMR spectra were obtained using a Bruker (400 MHz) NMR spectrophotometer in DMSO- d_6 . The chemical shifts were determined in parts per million (ppm) with respect to Tetramethylsilane (TMS).

Synthesis [27]. General procedure for Synthesis of azidic derivatives 2a and 2b.

One of the aniline compounds (15 mmol), (8-amino theophylline **2a**, 4-nitroaniline **2b**) was individually dissolved in 30 ml of a solution containing 10% hydrochloric acid (HCl). The solution was then cooled in an ice bath to 0 $^{\circ}\text{C}$, then 16.33 mmol of sodium nitrite (NaNO_2) dissolved in 5 ml of distilled water was added. The mixture was stirred for an hour at 0 degrees Celsius. Then 20 mmol of sodium azide dissolved in 5 ml of distilled water was added. The reaction mixture was stirred for one to two hours and then checked by TLC to detect the completion of the reaction. After completion of the reaction, a solution was treated with

5%NaOH and then extracted with chloroform (3×15 ml). Next, the organic liquid obtained was dried with anhydrous magnesium sulfate (MgSO_4) and then concentrated into a solid by a vacuum device. The resulting compounds were purified on a short column of SiO_2 using methanol-chloroform as eluent.

Com. 2a: Yield 83% , mp.62-64 $^{\circ}\text{C}$. IR, v , cm^{-1} : 3414 (NH), 2951, 2850 (C-H), 2136 (N_3), 1697,1674 (C=O), 1570, 1527 (C=C). ^1H NMR (DMSO- d_6 , ppm) δ : 6.60 s (1H, N-H), 5.83 s (1H, C₈-H), 4.62 s (1H, C₄-H), 3.20 s (3H, C₁₀-H), 2.78 s (3H, C₁₁-H). ^{13}C NMR (DMSO- d_6 , ppm) δ : 167.68 C-6, 159.63 C-3, 151.65 C-1, 79.66 C-8, 53.97 C-4, 36.38 C-11, 28.08 C-10. Anal. calc. for $\text{C}_7\text{H}_9\text{N}_7\text{O}_2$ (223.20 g/mol) : C, 37.67; H, 4.06; N, 43.93 Found: C, 36.03; H, 3.92; N, 43.25.

Com. 2b: Yield 81%; m.p. 67-69 $^{\circ}\text{C}$. IR, v , cm^{-1} : 3070 (C-H), 2120 (N_3), 1593 (C=C), 1516, 1346 (N-O). ^1H NMR (DMSO- d_6 , ppm) δ : 8.23 d ($J = 8.6$ Hz, 2H, C(3+5)-H), 7.33 d ($J = 8.7$ Hz, 2H, C(2+6)-H). ^{13}C NMR (DMSO- d_6 , ppm) δ : 147.13 C-4, 144.48 C-1, 125.98 C-(3+5), 120.51 C-(2+6). Analytical calculation for $\text{C}_6\text{H}_4\text{N}_4\text{O}_2$ (164.12 g/mol): C, 43.91; H, 2.46; N, 34.14. Found: C, 44.92; H, 2.46; N, 33.53.

Synthesis of 7-(2-chloroacetyl)-1,3-dimethyl-3,4,5,7-tetrahydro-1H-purine-2,6-dione 4. In a round flask, theophylline **3** (2.77 mmol, 0.5 gm) was dissolved in 25 ml of DMF, and Et_3N (2.77 mmol) was added to the solution at 10 $^{\circ}\text{C}$ and stirred for 30 minutes. Chloroacetyl chloride with an equivalent amount of moles was added gradually and slowly to the solution and stirred for an hour at room temperature; after that, the mixture was stirred at 60 $^{\circ}\text{C}$ and then followed by TLC. After completion of the reaction, a solution was treated with 5% NaOH and then extracted with chloroform. Next, the

organic layer was dried with anhydrous magnesium sulfate and then concentrated into a solid using a vacuum. The resulting compound was purified on a short column of SiO₂ using methanol–chloroform as eluent. Yield: 76%; m.p. 130–132 °C. IR, ν , cm⁻¹: 2974 (C-H), 1705, 1666 (C=O), 1566 (C=N). ¹H NMR (DMSO-*d*₆, ppm) δ : 8.81 s (1H, C₆-H), 6.71 s (1H, C₄-H), 5.76 s (1H, C₈-H), 3.86 s (2H, C₁₃-H), 3.35 s (3H C₁₀-H), 3.02 s (3H). ¹³C NMR (DMSO-*d*₆, ppm) δ : 168.60 C₁₄, 166.47 C₃, 154.86 C₁, 70.38 C₈, 55.76 C₄, 45.99 C₁₃, 34.56 C₁₁, 28.25 C₁₀. Analytical calculation for C₉H₁₁ClN₄O₃ (258.66 g/mol): C, 41.79; H, 4.29; N, 21.66. Found: C, 41.23; H, 4.16; N, 20.91.

Synthesis of 1,3-dimethyl-7-(2-(piperazin-1-yl)acetyl)-3,4,5,7-tetrahydro-1H-purine-2,6-dione 5. In a round flask, compound 4 (5 mmol) dissolved in 25 ml of DMF. Et₃N and piperazine are added to it with an equivalent amount of moles, and the mixture is stirred at 70 °C for a 3 hour (check by TLC). After completion of the reaction, a solution was treated with 5% NaOH and then extracted with chloroform. Next, the organic layer was dried with anhydrous magnesium sulfate (MgSO₄) and then concentrated into a solid using a vacuum. The resulting compound was purified on a short column of SiO₂ using methanol–chloroform as eluent. Yield: 72%; m.p. 185–187 °C. IR, ν , cm⁻¹, 3200 (NH), 2978, 2850 (C-H), 1708, 1662 (C=O), 1566 (C=N). ¹H NMR (DMSO-*d*₆, ppm) δ : 8.05 s (1H, C₆-H), 5.32 s (1H, C₄-H), 4.91 s (1H, C₈-H), 3.61 s (2H, C₁₃-H), 3.45 s (3H, C₁₀-H), 3.39 s (3H, C₁₁-H), 3.17 – 3.03 m (2H C_(15,16,18,19)-H), 2.54 s (1H, NH). ¹³C NMR (DMSO-*d*₆, ppm) δ : 161.70 C₁₂, 161.64 C₃, 154.88 C₁, 148.39 C₆, 82.60 C₈, 66.39 C₁₃, 55.79 C₄, 50.22 C₍₁₅₊₁₉₎, 45.96 C₁₆, 45.89 C₁₈, 34.67 C₁₁, 28.20 C₁₀. Analytical calculation for C₁₃H₂₀N₆O₃ (308.34 g/mol): C, 50.64; H, 6.54; N, 27.26. Found: C, 49.99; H, 6.17; N, 27.30.

Synthesis of 1,3-dimethyl-7-(2-(4-(prop-2-yn-1-yl)piperazin-1-yl)acetyl)-3,4,5,7-tetrahydro-1H-purine-2,6-dione 6. A mixture of compound 5 (5 mmol, 0.583 gm) with propargyl bromide (6 mmol, 1.111 gm) was dissolved in DMF (25 ml) and then stirred the reaction mixture for 16 h at 80 °C and monitored by thin layer chromatography (TLC). After the reaction was over, the mixture was

cooled and then poured onto ice flakes. Then the organic layer was separated from the aqueous layer using chloroform; the excess solvent was removed by distillation under vacuum pressure to obtain a white precipitate after purification with absolute ethanol. Yield: 79%; m.p. 169–171 °C. IR, ν , cm⁻¹, 3242 (C-H)_{alkyne}, 2978, 2850 (C-H)_{aliphatic}, 2210 (C $\equiv$ C), 1712, 1664 (C=O), 1566 (C=N). ¹H NMR (DMSO-*d*₆, ppm) δ : 8.07–7.93 m (1H, C₆-H), 5.18 s (1H, C₄-H), 4.52 s (1H, C₈-H), 3.64 d (*J* = 4.8 Hz, 1H, C₂₀-H), 3.42–3.35 m (5H, C₍₁₃₊₁₀₎-H), 3.24 s (3H, C₁₁-H), 2.68 s (1H, C₂₂-H), 2.47 t (*J* = 5.0 Hz, 4H), 2.41 t (*J* = 5.3 Hz, 4H). ¹³C NMR (DMSO-*d*₆, ppm) δ : 161.73 C₁₂, 161.27 C₃, 154.86 C₁, 148.31 C₆, 83.94 C₂₁, 79.19 C₈, 72.34 C₂₂, 53.60 C₁₃, 53.51 C₄, 52.10 C₍₁₅₊₁₉₎, 50.91 C₍₁₆₊₁₈₎, 48.12 C₂₀, 36.30 C₁₁, 28.19 C₁₀. Analytical calculation for C₆H₄N₄O₂ (346.39 g/mol): C, 55.48; H, 6.40; N, 24.26. Found: C, 55.23; H, 6.27; N, 24.09.

General procedure synthesis of 1,2,3-triazole derivatives 7 and 8. A mixture of compound 6 (2 mmol) with one of the azide derivatives 2a or 2b (2 mmol) was dissolved in 15 ml of DMF. After half an hour, anhydrous copper sulfate (0.170 mmol) and sodium ascorbate (0.170 mmol) were added to the reaction mixture, and then it was refluxed at a temperature of 60 °C for 8–10 hours. After the finish of the reaction (checked by TLC), the reaction mixture was cooled with ice, and the precipitate obtained was filtered and purified with absolute ethanol.

Com.7: Yield: 74%, mp. 202–204 °C. IR, ν , cm⁻¹, 3396 (N-H), 2968, 2873 (C-H)_{ali.}, 1708, 1647 (C=O), 1570 (C=N), 1539 (C=C), 1375 (C-N). ¹H NMR (DMSO-*d*₆, ppm) δ : 8.02 d (*J* = 29.6 Hz, 2H, C₍₆₊₂₂₎-H), 7.18 s (1H, NH), 6.12 s (1H, C₄-H), 5.40 s (2H, C₍₂₈₊₈₎-H), 4.10 (s, 1H, C₂₉-H), 3.74 (s, 2H, C₂₀-H), 3.40–3.37 m (8H, C₍₁₃₊₃₅₊₁₀₎-H), 3.23 s (3H, C₁₁-H), 2.95 s (3H, C₃₄-H), 2.44 m (4H, C₍₁₅₊₁₉₎-H), 2.37 m (4H C₍₁₆₊₁₈₎-H). ¹³C NMR (DMSO-*d*₆, ppm) δ : 161.75 C₁₂, 161.68 C₃₃, 161.29 C₃, 155.02 C₃₁, 151.66 C₁, 148.67 C₆, 138.73 C₋₂₆, 126.23 C₂₁, 123.37 C₂₂, 95.89 C₈, 70.82 C₂₉, 66.77 C₂₈, 57.16 C₄, 56.51 C₂₀, 54.65 C₁₃, 53.75 C₍₁₅₊₁₉₎, 53.00 C₍₁₆₊₁₈₎, 39.25 C₁₁, 30.19 C₁₀, 28.20 C₃₄, 27.95 C₃₅. Analytical calculation for C₂₃H₃₁N₁₃O₅ (569.59 g/mol): C, 48.50; H, 5.49; N, 31.97. Found: C, 47.88; H, 5.36; N, 31.76.

Com.8: Yield: 78, mp197-199°C. IR, ν , cm^{-1} , 3078 (C-H), 2926, 2875 (C-H)ali., 1797, 1651 (C=O), 1599 (C=N), 1547 (C=C), 1504, 1396 (N-O), 1303 (C-N). ^1H NMR (DMSO- d_6 , ppm) δ : 8.38-7.42 m (6H, $\text{C}_{\text{arom.}}$ -H), 5.80 s (1H, C4-H), 4.67 s (1H, C8-H), 3.71 s (1H, C20-H), 3.20 s (1H, C13-H), 3.14-3.11 s (6H, $\text{C}_{(11+10)}$), 2.36 s (4H, $\text{C}_{(16+18)}$ -H), 2.01 s (4H, $\text{C}_{(15+19)}$ -H). ^{13}C NMR (DMSO- d_6 , ppm) δ : 161.75 C_{12} , 161.67 C_{13} , 156.18 C_1 , 150.17 C_6 , 146.96 C_{29} , 142.19 C_{21} , 136.00 C_{26} , 126.85 $\text{C}_{(28+30)}$, 121.68 C_{22} , 119.83 $\text{C}_{(27+31)}$, 96.04 C_8 , 58.11 C_{13} , 57.65 C_4 , 55.15 $\text{C}_{(16+18)}$, 54.23 $\text{C}_{(15+19)}$, 45.95 C_{20} , 39.27 C_{11} , 28.12 C_{10} . Analytical calculation for $\text{C}_{22}\text{H}_{26}\text{N}_{10}\text{O}_5$ (569.59 g/mol): C, 51.76; H, 5.13; N, 27.44. Found: C, 51.12; H, 5.00; N, 27.28.

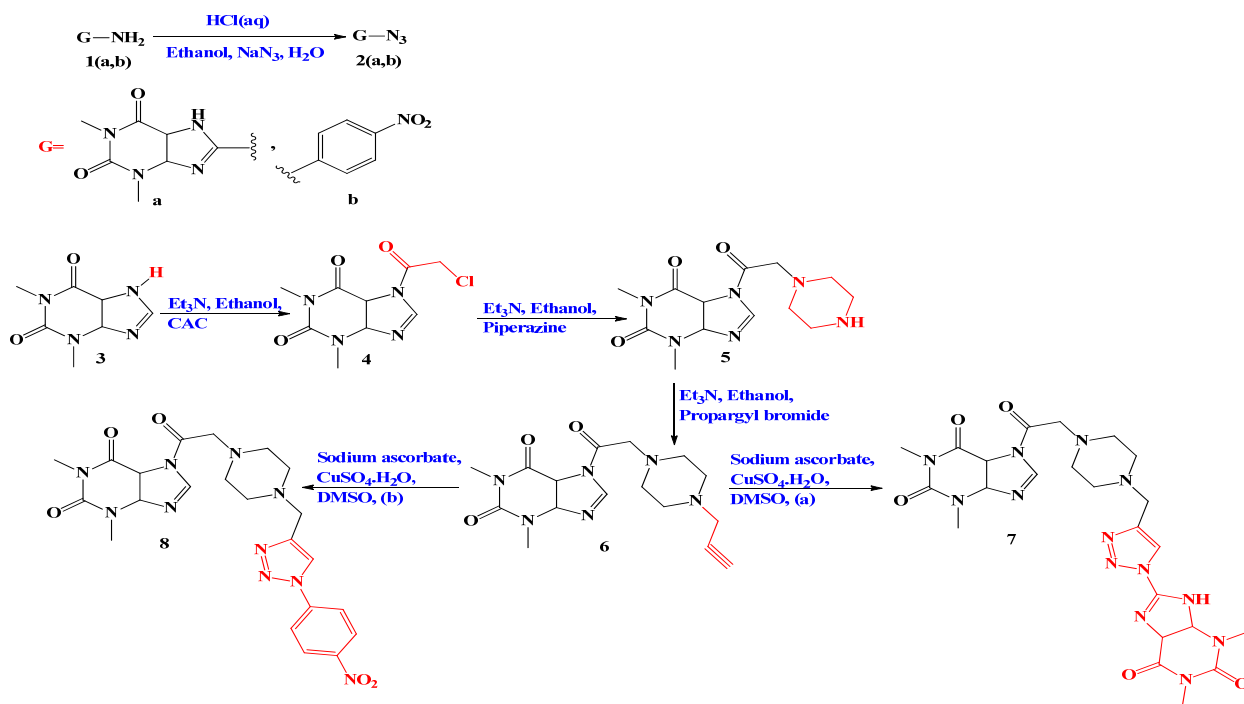
Antibacterial assay [28]. The following bacterial strains (*Staphylococcus aureus* and *Escherichia coli*) were employed in this study. The sensitivity of the bacteria to the compounds under investigation was ascertained using the well diffusion method. This method involved

preparing the culture medium per the supplier's instructions using 100 μL of Muller-Hinton medium (70191-Merk-Germany). After that, the medium was put on plates and given time to solidify. Using a cork drill, five depressions with a diameter of 6 mm were created in the medium after solidification. The inhibition zones had to be kept apart from one another by leaving enough space between each well. Thereafter, 0.1 mL of the bacterial suspension was transferred to the nutrient agar medium and evenly spread across the plate surface using a sterile cotton swab in a laminar flow hood. After that, the contaminated plates were stored for roughly 30 min. Each well received a volume of 0.1 mL that contained 100 μM and 250 μM of the compounds that were tested for biological activity. Each neighboring well received a different concentration, while one well was used as a control and received the same volume of solution.

Results and discussion

The derivatives studied in this manuscript were synthesized in sequential steps, where two types of azide derivatives, **2a** and **2b**, were prepared using the diazonium salt of the

corresponding amines. The derivative **4** was synthesized by reacting theophylline **3** with chloroacetyl chloride using Et_3N as a catalyst.



Scheme 1. The experimental steps for synthesis of compounds 2-8

After that, the derivative **5** was synthesized via a reaction of derivative **4** with piperazine in the presence of Et₃N as a catalyst. Next, derivative **5** was reacted with propargyl to synthesize and sodium ascorbate as catalysts. **Scheme 1** indicates the steps of the formation of these derivatives in sequence. In addition, structures of synthesized molecules were determined by

derivative **6** using Et₃N as a catalyst. After that, azides **2a** and **2b** were introduced individually in a cycloaddition reaction with the derivative **6** using both hydrated copper sulfate different techniques (FT-IR, ¹H NMR, and ¹³C NMR). The spectral data were included in the experimental section.

Table 1. Inhibition zones of antimicrobial activity for synthesized compounds in mm.

Antimicrobial activity				
<i>E. coli</i> (Gram-negative)		<i>S. aureus</i> (Gram-positive)		
No.	Size mm		Size mm	
	100 µM	250 µM	100 µM	250 µM
4	20	20	16	22
5	10	14	10	15
6	16	20	18	25
7	16	26	13	20
8	14	22	15	23
Cephalexin	22	26	12	15
Penicillin	11	15	20	25

Antibacterial Activity. Control of microbial populations is essential to prevent the emergence of diseases, infections, decomposition, spoilage, and contamination that they cause. Newly synthesized compounds were examined for their antimicrobial activity *in vitro* against bacteria (*Staphylococcus aureus* and *Escherichia coli*). Results of antimicrobial

activity revealed that most of the tested compounds had moderate to strong activity. Compounds were appearing to have a strong effect against *Escherichia coli*, except derivative **5** was appearing to have a middling effect. In addition, compounds had a middling effect against *Staphylococcus aureus* compared to Cephalexin, as illustrated in Table 1.

Conclusion

Theophylline derivatives were synthesized in yields ranging from 72 to 82%. Some of the synthesized compounds exhibited encouraging antibacterial properties compared to that of standard reference, and this may be due to their structural properties that help in making

morphological defects in the cell walls of bacteria. Fourier transform infrared spectroscopy (FT-IR), proton nuclear magnetic resonance ¹H NMR and ¹³C NMR techniques were used to validate the chemical structures of the synthesized compounds.

Acknowledgments

The authors would like to express their gratitude to the Department of Chemistry, College of Education, University of Al-Qadisiyah for their excellent assistance in conducting this work. The authors would also like to express their gratitude to the University of Basrah for their valuable support in conducting NMR spectroscopy measurements.

Conflict of interest

The authors declare that there is no conflict of interests regarding the publication of this manuscript.

References

1. Nabeel A.A.R., Islam H.T. Synthesis Characterization *in Silico* and *in Vitro* Study of New 1, 2, 3-Triazole Derivatives as Antioxidant Agents. *Chemical Problems*, 2023, **Vol.** 21(4), p. 343-352. DOI:[10.32737/2221-8688-2023-4-343-352](https://doi.org/10.32737/2221-8688-2023-4-343-352)
2. Brüggemann R.J., Verheggen R., Boerrigter E., Stanzani M., Verweij P.E., Blijlevens N.M.A., Lewis R.E. Management of drug-drug interactions of targeted therapies for haematological malignancies and triazole antifungal drugs. *The Lancet. Haematology*, 2022, **Vol.** 9(1), p. e58–e72. DOI:[10.1016/S2352-3026\(21\)00232-5](https://doi.org/10.1016/S2352-3026(21)00232-5)
3. Abdul-Rida N.A., Sayyah M.H., Jaber Q.A.H. Synthesis, characterization, efficiency evaluation of some novel triazole derivatives as acid corrosion inhibitors. *Int. J. Corr. Scale Inhib.*, 2023, **Vol.** 12(1), p. 101-125. DOI:[10.17675/2305-6894-2023-12-1-6](https://doi.org/10.17675/2305-6894-2023-12-1-6)
4. Matin M.M., Matin P., Rahman M.R., Ben Hadda T., Almalki F.A., Mahmud S., Ghoneim M.M., Alruwaily M., Alshehri S. Triazoles and Their Derivatives: Chemistry, Synthesis, and Therapeutic Applications. *Frontiers in molecular biosciences*, 2022, **Vol.** 9, 864286. DOI:[10.3389/fmolb.2022.864286](https://doi.org/10.3389/fmolb.2022.864286)
5. Avula S.K., Khan M., Halim S.A., Khan A., Al-Riyami S.A., Csuk R., Das B., Al-Harrasi A. Synthesis of New 1H-1,2,3-Triazole Analogs in Aqueous Medium via "Click" Chemistry: A Novel Class of Potential Carbonic Anhydrase-II Inhibitors. *Frontiers in chemistry*, 2021, **Vol.** 9, 642614. DOI:[10.3389/fchem.2021.642614](https://doi.org/10.3389/fchem.2021.642614)
6. Khan M., Avula S.K., Halim S.A., Waqas M., Asmari M., Khan A., Al-Harrasi A. Biochemical and *in silico* inhibition of bovine and human carbonic anhydrase-II by 1H-1,2,3-triazole analogs. *Front. Chem.*, 2022, **Vol.** 10, 1072337. DOI:[10.3389/fchem.2022.1072337](https://doi.org/10.3389/fchem.2022.1072337)
7. Jaber Q.A., Shentaif A.H., Almajidi M., Ahmad I., Patel H., Azad A.K., Alnasser S.M., Alatawi H.A., Menaa F., Alfaiifi S.Y.M., Rahman M.M., Ali M.M., Aditya Rao S.J. Synthesis, Structure, and *In Vitro* Pharmacological Evaluation of some New Pyrimidine-2-Sulfonamide Derivatives and Their Molecular Docking Studies on Human Estrogen Receptor Alpha and CDK2/Cyclin Proteins. *Russ J Bioorg. Chem.*, 2023, **Vol.** 49, S106-S118. DOI:[10.1134/S1068162023080095](https://doi.org/10.1134/S1068162023080095)
8. Ewies E.F., Sabry E., Bekheit M.S., Fouad M.A., Vullo D., Supuran C.T. Click chemistry-based synthesis of new benzenesulfonamide derivatives bearing triazole ring as selective carbonic anhydrase II inhibitors. *Drug Dev. Res.*, 2022, **Vol.** 83(6), p. 1281–1291. DOI:[10.1002/ddr.21957](https://doi.org/10.1002/ddr.21957)
9. Buza A., Türkeş C., Arslan M., Demir Y., Dincer B., Nixha A.R., Beydemir Ş. Discovery of novel benzenesulfonamides incorporating 1,2,3-triazole scaffold as carbonic anhydrase I, II, IX, and XII inhibitors. *Int. J. Boil. Macromol.*, 2023, **Vol.** 239, 124232. DOI:[10.1016/j.ijbiomac.2023.124232](https://doi.org/10.1016/j.ijbiomac.2023.124232)
10. Avula S.K., Khan A., Rehman N.U., Anwar M.U., Al-Abri Z., Wadood A., Riaz M., Csuk R., Al-Harrasi A. (2018) Synthesis of 1H-1,2,3-triazole derivatives as new α -glucosidase inhibitors and their molecular docking studies. *Bioorg. Chem.*, 2018, **Vol.** 81, p. 98–106. DOI:[10.1016/j.bioorg.2018.08.008](https://doi.org/10.1016/j.bioorg.2018.08.008)
11. Olivelli A.G., Olelewe C., Wolff L., Parkin S., Webster C.E., Awuah S.G., Huckaba A. Stable Cu (I) Complexes for Intracellular Cu-Catalyzed Azide Alkyne Cycloaddition. *Chemistry (Weinheim an der Bergstrasse, Germany)*, 2024, **Vol.** 30(72), e202402887. DOI:[10.1002/chem.202402887](https://doi.org/10.1002/chem.202402887)
12. Hayallah A.M., Elgaher W.A., Salem O.I., Alim A.A. Design and synthesis of some new theophylline derivatives with bronchodilator and antibacterial activities. *Arch. Pharm. Res.*, 2011, **Vol.** 34(1), p. 3–21. DOI:[10.1007/s12272-011-0101-8](https://doi.org/10.1007/s12272-011-0101-8)
13. Jenne J.W. Theophylline as a bronchodilator in COPD and its combination with inhaled beta-adrenergic drugs. *Chest*, 1987, **Vol.** 92(1), p. 7S–14S.
14. Ashihara H., Yokota T., Crozier A. *Purine Alkaloids, Cytokinins, and Purine-Like Neurotoxin Alkaloids*. In: Ramawat K.,

- Mérillon J.M. (eds). 2013. Natural Products. Springer. Berlin. Heidelberg. DOI:[10.1007/978-3-642-22144-6_32](https://doi.org/10.1007/978-3-642-22144-6_32)
15. Torres-Borrego J., Sánchez-Solís M. Dissecting Airborne Allergens. *J. Clin. Med.*, 2023, **Vol. 12(18)**, 5856. DOI:[10.3390/jcm12185856](https://doi.org/10.3390/jcm12185856)
16. Scurek M., Brat K. A narrative review of theophylline: is there still a place for an old friend. *J. Thorac Dis*, 2024, **Vol. 16(5)**, p. 3450-3460. DOI:[10.21037/jtd-23-1781](https://doi.org/10.21037/jtd-23-1781)
17. Pardo M., Paul N.E., Collins-Praino L.E., Salamone J.D., Correa M. The non-selective adenosine antagonist theophylline reverses the effects of dopamine antagonism on tremor, motor activity and effort-based decision-making. *Pharmacol Biochem Behav.*, 2020, **Vol. 198**, 173035. DOI:[10.1016/j.pbb.2020.173035](https://doi.org/10.1016/j.pbb.2020.173035)
18. Barnes P.J. Theophylline. *Am J Respir Crit Care Med.*, 2013, **Vol. 188(8)**, p. 901-906. DOI:[10.1164/rccm.201302-0388PP](https://doi.org/10.1164/rccm.201302-0388PP)
19. Landells L.J., Jensen M.W., Orr L.M., Spina D., O'Connor B.J., Page C.P. The role of adenosine receptors in the action of theophylline on human peripheral blood mononuclear cells from healthy and asthmatic subjects. *Br J Pharmacol.*, 2000, **Vol. 129(6)**, p. 1140-1144. DOI:[10.1038/sj.bjp.0703177](https://doi.org/10.1038/sj.bjp.0703177)
20. Takanashi S., Hasegawa Y., Kanehira Y., Yamamoto K., Fujimoto K., Satoh K., Okamura K. Interleukin-10 level in sputum is reduced in bronchial asthma, COPD and in smokers. *The Eur. Respir. J.*, 1999, **Vol. 14(2)**, p. 309-314. DOI:[10.1034/j.1399-3003.1999.14b12.x](https://doi.org/10.1034/j.1399-3003.1999.14b12.x)
21. Saeed S., Shahzadi I., Zahoor A.F., Al-Mutairi A.A., Kamal S., Faisal S., Irfan A., Al-Hussain S.A., Muhammed M.T., Zaki M.E. Exploring theophylline-1,2,4-triazole tethered *N*-phenylacetamide derivatives as antimicrobial agents: unraveling mechanisms via structure-activity relationship, *in vitro* validation, and *in silico* insights. *Front. Chem.*, 2024, **Vol. 12**, 1372378. DOI:[10.3389/fchem.2024.1372378](https://doi.org/10.3389/fchem.2024.1372378)
22. Jia Y., Yang D., Wang W., Hu K., Yan M., Zhang L., Gao L., Lu Y. Recent advances in pharmaceutical cocrystals of theophylline. *Nat Prod Bioprospect.*, 2024, **Vol. 14(1)**, 53. DOI:[10.1007/s13659-024-00470-y](https://doi.org/10.1007/s13659-024-00470-y)
23. Montañó L.M., Sommer B., Gomez-Verjan J.C., Morales-Paoli G.S., Ramírez-Salinas G.L., Solís-Chagoyán H., Sanchez-Florentino Z.A., Calixto E., Pérez-Figueroa G.E., Carter R., Jaimez-Melgoza R., Romero-Martínez B.S., Flores-Soto E. Theophylline: Old Drug in a New Light, Application in COVID-19 through Computational Studies. *Int J Mol Sci.*, 2022, **Vol. 23(8)**, 4167. DOI:[10.3390/ijms23084167](https://doi.org/10.3390/ijms23084167)
24. Mangasuli S.N., Hosamani K.M., Devarajegowda H.C., Kurjogi M.M., Joshi S.D. Synthesis of coumarin-theophylline hybrids as a new class of anti-tubercular and anti-microbial agents. *Eur J Med Chem.*, 2018, **Vol. 146**, p. 747-756. DOI:[10.1016/j.ejmech.2018.01.025](https://doi.org/10.1016/j.ejmech.2018.01.025)
25. Chang Y.L., Hsu Y.J., Chen Y., Wang Y.W., Huang S.M. Theophylline exhibits anti-cancer activity *via* suppressing SRSF3 in cervical and breast cancer cell lines. *Oncotarget*. 2017, **Vol. 8(60)**, p. 101461-101474. DOI:[10.18632/oncotarget.21464](https://doi.org/10.18632/oncotarget.21464)
26. Eid N.S., O'Hagan A., Bickel S., Morton R., Jacobson S., Myers J.A. Anti-inflammatory dosing of theophylline in the treatment of status asthmaticus in children. *J Asthma Allergy*, 2016, **Vol. 9**, p. 183-189. DOI:[10.2147/JAA.S113747](https://doi.org/10.2147/JAA.S113747)
27. Abdul-Rida N.A., Tarrad I.H. Design, Synthesis, Molecular Docking Study, and Biological Evaluation of Some New 1,2,3-Triazole Derivatives as Anticancer and Antioxidant Agents. *Russ J Gen Chem.*, 2023, **Vol. 93**, p. 2874-2882. DOI:[10.1134/S107036322311018X](https://doi.org/10.1134/S107036322311018X)
28. Jaber Q.A.H., Abdul-Rida N.A., Adnan S. Boosting 3*H*-Benzo[*f*]chromen-3-one Chalcone with Anti-inflammatory Drugs: Synthesis, Characterization, and Evaluation of Cytotoxicity and Antimicrobial Activity. *Russ. J. Org. Chem.*, 2020, **Vol. 56**, p. 1622-1627. DOI:[10.1134/S1070428020090195](https://doi.org/10.1134/S1070428020090195)